18-DEOXYALDOSTERONE, ITS CHEMICAL AND MICROBIAL REDUCTION PRODUCTS

M. HARNIK,*† Y. AHARONOWITZ‡ and R. LAMED[†] The George S. Wise Faculty of Life Sciences, Tel Aviv University, Tel Aviv, 69978, Israel

(Received in the U.K. 26 March 1982)

Abstract—Reduction of the diketal 1 with sodium aluminum bis-(methoxyethoxy) hydride afforded the crystalline 18-hydroxycorticosterone diketal (2), an intermediate in the formation of 18-deoxyaldosterone acetate (4b). The hitherto unreported but anticipated metabolites of 4 were prepared as follows: hydrogenations of 4b furnished the 5α - and 5β isomers 6 and 5b, and thence the tetra- and hexahydro derivatives 10, 11, 8, 9 and 7, and the 3-deoxy compounds 12 and 13. Anaerobic fermentations of 4b with *Clostridium paraputrificum* gave the tetrahydro derivative 8b in high yield.

18-Deoxyaldosterone (18-DAL) (4a) is easily prepared by dehydration of 18-hydroxycorticosterone, a naturally occurring corticoid; it has been suggested that 4a is identical with the "less polar" form of the latter, and may be formed in the organism by non-enzymatic dehydration.¹ 18-DAL has been shown to possess a high affinity for mineralocorticoid receptors and is a partial agonist/predominant antagonist to aldosterone in rat and toad mineralocorticoid test systems. Because of this low affinity for androgen receptors it may offer advantages over spironolactone, a therapeutic agent used in the treatment of hypertension, whose main drawback is associated with estrogenic effects.^{2,3}

Since di- and tetrahydro metabolites of aldosterone exhibit mineralocorticoid activity,⁴ we decided to synthesize several reduced derivatives of 18-DAL for biological testing, in order to find out if, by analogy, the antimineralocorticoid properties of 18-DAL are retained in them; this objective could now be reached by the employment of chemical and microbiological methods.

18-DAL had previously been synthesized in several ways. In the first method (Scheme 1), LAH reduction of the racemic lactone 1 furnished the 18-hydroxycortiscosterone derivative 2, which on boiling in 90% acetic acid afforded racemic $4.^{5-7}$ 18-Hydroxycorticosterone itself was converted into 4a by *p*-toluenesulfonic acid in boiling ethylene dichloride.² In the second method, microbial hydroxylation of corticosterone with *Cory-nespora cassiicola* yielded the dimer of 18-hydroxy-corticosterone (3), which, following treatment with boiling acetic acid, gave mostly the acetate 4b.⁸ In the third method, 1-dehydrocorticosterone acetate was converted into its iodo derivative via the 11-nitrite and then directly transformed into 1-dehydro-18-DAL acetate, which on reduction with Wilkinson's catalyst gave 4b.⁹

While the last route is the shortest, the first is the method of choice when the lactone or its diketal 1 is readily available. We have now found that this approach can be simplified if sodium aluminum bis-(methoxyethoxy) hydride (SAMH) is used instead of LAH in the reduction of 1 to afford the crystalline 3,20-diketal of 18-hydroxycorticosterone (2) in high yield. Undoubtedly other complex hydrides soluble in organic solvents can also be used. Heating the ketal 2 in methylene dichloride (MDC) containing p-toluenesulfonic acid furnished predominantly the dimer 3, with some free 18-DAL (4a). The dimer was converted into 4a with hot acetic acid, essentially as previously described,⁸ and then acetylated. With 4b readily available, we set out to prepare several hitherto unreported reduced derivatives.

For conversion into the di- and tetrahydro derivatives, 4b was hydrogenated with Pd to the 5α -dihydro compound 6. The 5 β -epimer 5b could be isolated from the filtrate in the crystalline form only after seeding with material obtained by the microbial route described below. Catalytic reduction of 6 with raney Ni in dioxane gave the hexahydro compound 7, and not the expected^{10,11} tetrahydro derivative 10b. Verification of structure was obtained by treatment of 7 with the Jones reagent which caused reversal of the reaction with formation of the dione 6. Alternatively, hydrogenation of 6 with Pd in ethanol in the presence of acetic acid for 2 days at 30 psi yielded the 3β -ol 10b, in analogy with related conversions of other 3-ketosteroids under these conditions.^{12,13} Hydrolysis of the acetate group in position 21 was performed with hydrochloric acid in a MDCmethanol mixture to give the free diol 10a, having a broad m.p. However, acetylation of this product furnished the sharp-melting diacetate 10c. No appreciable epimerization at position 17 occurred during the acid hydrolysis of 10b, as was shown by acetylation of the monoacetate 10b, whereupon the diacetate 10c was obtained, identical with the sample obtained by acetylation of 10a. This proof for retention of configuration was desirable since extensive inversion of 18-DAL acetate (4b) at position 17 can easily occur.9

Inversion of configuration at position 3 was effected by treatment of 10b with *p*-toluenesulfonyl chloride in pyridine to afford the tosylate 10d, followed by solvolysis in dimethylformamide. Two new compounds were isolated: the elimination product 12, which could be hydrogenated with Pd in ethyl acetate to the saturated ester 13, and the 3α -formate 11b. Acidic hydrolysis of the latter, under conditions identical with those used for 10b, yielded the diol 11a; this on acetylation afforded the diacetate 11c.

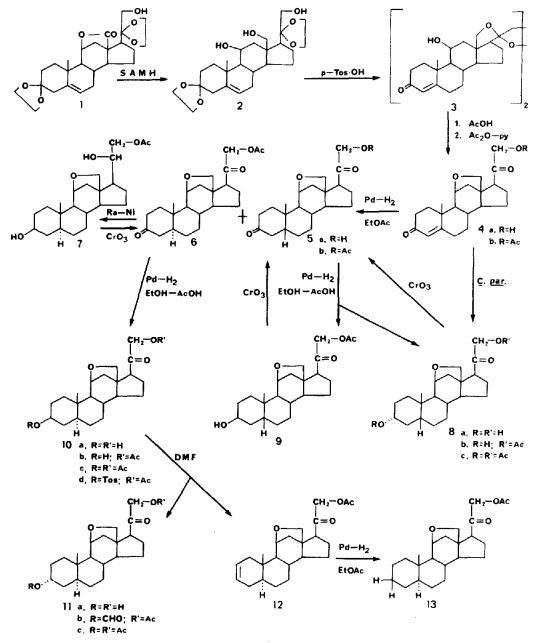
^{*}To whom inquiries should be addressed.

[†]Center for Biotechnology.

[‡]Department of Microbiology.

Since the 5β -dihydro siomer **5b** could not be obtained in crystalline form by the Pd in ethyl acetate hydrogenation of **4b**, the application of a microbial bioconversion step was also investigated and found to be extremely useful in solving this synthetic problem. Anaerobic sterospecific reductions with *Clostridium* paraputrificum to afford the 5β di- and $3\alpha,5\beta$ -tetrahydro derivatives had been successfully applied to a variety of Δ^4 -3-ketosteroids,^{14b} although aldosterone could not be satisfactorily reduced by this method.¹⁵ We have now found that incubation of 18-DAL acetate **4b** for 24 h with this organism furnished the desired $3\alpha,5\beta$ -tetrahydroester **8b** in high yield. In contrast to a corresponding reduction of 18-hydroxy-11-deoxycorticosterone 21acetate by this organism, which gave the free 21hydroxy- 3α , 5β -tetrahydro derivative, $^{14\alpha}$ there was no hydrolytic cleavage of the acetate during the fermentation of 4b. Acetylation of 8b furnished the diacetate 8c, and acidic hydrolysis of the latter gave the diol 8a; as in the case of 10b, no detectable inversion took place at position 17, since acetylation of this diol gave a product identical with 8c. Jones oxidation of 8b furnished the 3-one 5b, identical with the material accompanying the 5α -isomer 6 in the mild Pd hydrogenation of 4b. Acidic hydrolysis of 5b yielded the 21-ol 5a, and reacetylation of 5a regenerated the starting acetate 5b, showing again that the 17β -configuration was preserved.

As in the case of the 5α -isomer 6, prolonged hydrogenation of 5b in ethanol containing some acetic acid at 20 psi in the presence of Pd caused reduction of the



Scheme 1.

ketone function at position 3, and the two epimeric 3α and 3β -ols **8b** and **9**, obtained in the ratio 1:10, were separated by chromatography. As with **8b**, **9** could be reoxidized to **5b** with the Jones reagent, confirming the relationship between these 3 compounds.

EXPERIMENTAL

Merk A.G. silica gel 60 was employed in all column chromatograms. NMR spectra were obtained in $CDCl_3$ (TMS) on a Jeol 60 MHz spectrophotometer. IR spectra were taken on a Perkin-Elmer 297 spectrometer in KBr pellets. Mass spectra were recorded with a Dupont 21-491B spectrometer. TLC spots were visualized by spraying with 10% H₂SO₄ in EtOH prior to heating.

11,6,18,21 - Trihydroxy - 5 - pregnene - 3,20 - dione 3,20 di - (ethylene glycol) ketal (3,20 - diketal of 18 - hydroxycorticosterone) (2). A suspension of 9.00 g of 11,6,21 - dihydroxy - 5 pregnene - 3,20 - dione - 18 - oic acid 3,20-di-(ethylene glycol) ketal (18 \rightarrow 11) lactone (1)⁵ in 150 ml dry benzene was cautiously treated with 20 ml of 70% soln of SAMH in benzene. After the initial vigorous reaction subsided, the clear soln was refluxed for 1 h, cooled, and then cautiously treated with swirling with a total of 200 ml 10% NaOHaq. The mixture was diluted with 300 ml MDC and magnetic stirring applied. After 15 min the ppt was collected and washed with water to afford 6.20 g of 2, m.p. 191-197° (sinter at 173°). The aqueous phase was extracted twice with MDC, the extracts washed with 10% NaClaq., dried with Na₂SO₄, treated with 0.5 ml Et₃N and concentrated in vacuo to furnish 2.67 g of the same material, m.p. 185-190°. Further concentration gave 0.30 g, m.p. 155-185°.

18-Deoxyaldosterone acetate (4b). A suspension of 9.8 g of 2 in 500 ml MDC was treated with 1.05 g p-toluenesulfonic acid monohydrate and the mixture was refluxed for 40 min. A few min after the solid dissolved the crystalline dimer 3 began to precipitate. The mixture was cooled in ice, treated with 200 ml sat. NaHCO3aq., and mechanically stirred for 15 min. The dimer 3 was collected and washed with water to afford 3.53 g, m.p. 295-303° (reported⁸ 293-296°). The organic layer, containing mostly a mixture of 3 and 4a, was taken to dryness, combined with the solid 3 isolated above and refluxed in 250 ml AcOH for 80 min, during which time the solid gradually dissolved. The solvent was removed in vacuo, the crude product acetylated overnight at room temp. with 60 ml each of Ac₇O and pyridine, and worked up by adding ice and water up to a volume of 11., extracting with MDC, washing with dil. HCl and finally with NaHCO3aq. Evaporation of solvent afforded the crude 18-DAL acetate (4b), m.p. 150-162°, which was then chromatographed. Elution with 20% acetone in petroleum ether followed by crystallization from MDC-ether furnished 5.26 g of material, m.p. 165-168° (reported 160-161°;8 161-163°9).

Hydrogenation of 4b. A soln of 2.057 g of 4b in 150 ml EtOAc was hydrogenated for 2 h in the presence of 0.23 g of 5% Pd-C. The filtered soln was evaporated in vacuo and the solid was washed with cold ether to afford 1.451 g of 21 - hydroxy - 11 β ,18 - oxido - 5 α - pregnane - 3,20 - dione acetate (6). The pure sample (MDC-ether) had the m.p. 191-193°; λ_{max}^{RB} 5.72, 5.78 and 5.85 μ ; δ 1.09 (s, 19-CH₃), 2.16 (s, 21-OAC), 2.97 (t, $J = 7.9, 17\alpha$ -H), 3.37 and 3.64 (ABq, J = 8.8, 18-CH₂), 4.43 (d, $J = 6.4, 11\alpha$ -H), 4.62 (s, 21-CH₂) ppm; m/e 388. The ether filtrate was seeded with 21 - hydroxy - 11 β ,18 - oxido - 5 β - pregnane - 3,20 - dione acetate (5b) to deposit 0.115 g of this material of m.p. 95-100°. Chromatography of the filtrate (elution with 25% acetone in petroleum ether), identical with the material described below.

3 $\beta_20\beta_21$ - Trihydroxy - 11 β_1 18 - oxido - 5 α - pregnane 21 acetate (7). A soln of 202 mg of 6 in 15 ml dry dioxane was hydrogenated in the presence of 500 mg raney Ni for 2 h at atmospheric pressure. The filtered soln was evaporated to dryness and the residue triturated with ether to yield 76 mg of 7, m.p. 177-191°. The pure sample melted at 195-198° (EtOAc) and displayed in TLC a blue coloration with ethanolic H₂SO₄; $\lambda_{\text{KBr}}^{\text{KBr}}$ 5.86 μ ; δ 0.87 (s, 19-CH₃), 2.06 (s, 21-OAc), ca 3.80 (broad m, 3 α -H, 17 α -H), 3.95 and 4.07 (ABq, 21-CH₂), 4.32 (d, J = 6.5, 11 α -H) ppm; m/e 392. Oxidation of 7 to 6. A cooled soln of 20 mg of 7 in 12 ml acetone was treated with 0.1 ml Jones reagent. After 5 min at 5° 0.5 ml MeOH was added, and after a further 5 min, 10 ml water. The clear soln was concentrated *in vacuo* at room temp. to remove the acetone. The colorless crystals were collected and washed with water to furnish 15 mg of material, m.p. 186-191°, whose IR spectrum and TLC were identical with those of 6 obtained above.

3 β ,21 - Dihydroxy - 11 β ,18 - oxido - 5 α - pregnane - 20 - one 21acetate (10b). A hot soln of 1.45 g of 6 in 250 ml EtOH was quickly cooled to room temp., 2 ml AcOH and 1.6 g 5% PdC was added, and the mixture was hydrogenated at room temp. at 30 psi for 44 h in a Parr apparatus. The filtered soln was evaporated in vacuo and the oily residue crystallized by dissolving in 30 ml ether and chilling, to give 0.95 g of 10b, m.p. 154-158°. A second crop of 0.25 g of slightly less pure material was obtained by concentration, and some more by chromatography, eluting with 20% acetone in petroleum ether. The pure sample melted at 158-159.5° (ether); λ_{max}^{RB7} 2.83, 5.70 and 5.80 μ ; δ 0.89 (s, 19-CH₃), 2.14 (s, 21-OAc), 3.60 (m, 3 α -H), 3.01 (t, J = 8, 17 α -H), 3.34 and 3.54 (ABq, J = 8.2, 18-CH₂), 4.33 (d, J = 6.5, 11 α -H), 4.55 (s, 21-CH₂) ppm; m/e 390.

3 β_{1} - Dihydroxy - 11 β_{1} 18 - oxido - 5 α - pregnane - 20 - one 3 - tosylate 21 - acetate (10d). A soin of 0.91 g of the 3 β -ol 10b in 15 ml pyridine was treated with 4 g p-toluenesulfonyl chloride and the mixture was magnetically stirred for 19 h at room temp. Additional 4 g p-toluenesulfonyl chloride was then added and stirring was continued for additional 24 h. Ice and water were added up to a volume of 110 ml and the mixture was stirred for 1 h at 0°. The crystals were collected and washed with water to furnish 1.23 g of 10d, about 85% pure, which after recrystallization from ether melted at 162-163° (dec); λ_{max}^{RBF} 5.71, 5.80 and 6.26 μ .

21 - Hydroxy - 11 β , 18 - oxido - 5α - pregn - 1 - ene - 20 - one acetate (12) and 3α , 21 - dihydroxy - 11 β , 18 - oxido - 5α pregnane - 20 - one 3-formate 21 - acetate (11b). A soln of 1.23 g of crude 10d in 15 ml DMF was kept at 80° for 68 h, whereupon the solvent was distilled *in vacuo*, and ice and water were added up to a volume of 20 ml. The soft solid was collected, washed with water and chromatographed. Elution with 20% acetone in petroleum ether gave first the Δ^2 compound 12, which after recrystallization from MeOH weighed 0.22 g and melted at 129-131°; a further recrystallization gave the m.p. 131-132°; λ_{max}^{max} 5.70 and 5.83 μ ; δ 0.84 (s, 19-CH₃), 2.15 (s, 21-OAc), 3.01 (t, J = 8, 17α -H), 3.28 and 3.58 (ABq, J = 8.2, 18-CH₂), 4.30 (d, J = 6.5, 11α -H), 4.52 (s, 21-CH₂), 5.58 (2H s, 2-H₃-H) pm; *m/e* 372.

Further elution afforded 11b, which after recrystallization from MeOH weighed 0.44 g, m.p. 180-191° (single TLC spot), narrowed by another recrystallization to 185-186°; $\lambda_{max}^{KBr} 5.72-5.82 \mu$; m/e 418.

Continued elution with 20% acetone in petroleum ether afforded 0.04 g of the starting tosylate 10d, m.p. 158-159° (dec). A repeat chromatography of the combined filtrates effected

separation of further amounts of 12, 11b and 10d.

 $3\alpha_2 21 - Dihydroxy - 11\beta_1 8 - oxido - 5\alpha - pregnane - 20 - one$ (11a). A soln of 50 mg of 11b in 1.3 ml MDC and 4.3 ml MeOH was treated with a mixture of 0.6 ml conc HCl and 0.33 ml water, and allowed to stand at room temp. for 3 days. The soln was diluted with MDC and extracted with sat. NaHCO₃aq. The aqueous phase was backwashed with MDC, the combined MDC extracts were evaporated and the residue was treated with ether to afford 17 mg, m.p. 183-187°. Recrystallization from MDC yielded the diol 11a of m.p. 192-196° (dec); $\lambda_{max}^{\rm KBr} 5.84 \mu$; m/e 348.

 $3\alpha_2 1 - Dihydroxy - 11\beta_1 8 - oxido - 5\alpha - pregname - 20 - one diacetate (11c). A 79 mg sample of 11a of m.p. 174-181° (about 80% pure) was treated overnight with 1 ml each of pyridine and Ac₂O. Addition of 15 ml ice and water afforded a solid which was collected, washed with water and recrystallized from MeOH to furnish 48 mg of 11c, m.p. 172-175°. The pure sample melted at 175-177°; <math>\lambda_{max}^{RBT} 5.72-5.80 \mu$; $\delta 0.87$ (s, 19-CH₃), 2.01 (s, 3-OAc), 2.13 (s, 21-OAc), 3.02 (t, J = 8, 17 α -H), 3.35 and 3.57 (ABq, J = 8, 18-CH₂), 4.39 (d, J = 6.5, 11 α -H), 4.59 (s, 21-CH₂), 5.00 (t, 3 β -H) ppm; mle 372 (M-60).

 $3\beta_2 1 - Dihydroxy - 11\beta_1 8 - oxido - 5\alpha - pregnane - 20 - one (10a). A soln of 85 mg of 10b in 2.2 ml MDC and 7 ml MeOH was$

treated with a mixture of 1 ml conc. HCl and 0.55 ml water. After letting stand for 3 days at room temp. the mixture was worked up as described above for the preparation of 11a. The ether-washed product (45 mg) was homogeneous in TLC but displayed a wide m.p. $(140-158^\circ)$, not much changed on recrystallization from MDC-petroleum ether.

 3β ,21 - Dihydroxy - 11 β ,18 - oxido - 5α - pregnane - 20 - one diacetate **10c**

(a) From the diol 10a. A soln of 35 mg of diol in 0.4 ml of each pyridine and Ac_2O was stored overnight at room temp. Quenching with ice-water furnished the diacetate, over 90% pure (TLC), which after recrystallization from MeOH melted at 164–167°, and whose IR spectrum was identical with that of a sample of 10c described below.

(b) From the monoacetate 7a. Treatment of 151 mg of 10b with 1.5 ml each of pyridine and Ac₂O furnished 10c which was recrystallized from MeOH to yield 125 mg, m.p. 168-170°. The pure sample melted at 170-172°; λ_{max}^{RBT} 5.68, 7.72 and 5.78-5.81 μ ; δ 0.93 (s, 19-CH₃), 2.03 (s, 3-OAc), 2.18 (s, 21-OAc), 3.04 (t, J = 8, 17 α -H), 3.34 and 3.63 (ABq, J = 8, 18-CH₂), 4.38 (d, J = 6.5, 11 α -H), 4.60 (m, 3α -H), 4.62 (s, 21-CH₂) ppm; *mle* 372 (M-60).

Microbiological conversion of 18-DAL acetate (4b) into 3α ,21 dihydroxy - 11, 8, 18 - oxido - 5, B - pregnane - 20 - one 21 acetate (8b). A 200 ml narrow-mouth bottle was charged with 120 ml of a sterile nutrient soln prepared as described.^{14a} Finely powdered 18-DAL acetate (4b) (200 mg) was then added under sterile conditions under N2, and the mixture was anaerobically inoculated with a pregrown (without added steroids) culture of Clostridium paraputrificum ATCC 25780 and shaken in a gyratory shaker (250 rpm) at 37° for 24 h. At the end of that period the culture (pH 5.3-5.8) containing a voluminous white ppt was stirred with 50 ml of MDC for 15 min, the mixture was filtered with suction through a celite pad, and the aqueous phase was twice reextracted with 30 ml MDC. The combined extracts were washed with NaHCO3aq., dried and evaporated to a gum which crystallized on contact with ether to afford 155 mg of 8b, having a double m.p. 100-105° and 125-130°. A further 20 mg was obtained by chromatography of the filtrate. The pure material melted at 134-135° (EtOAc-petroleum ether); λ KBr 5.71 and 5.75 μ; δ 0.97 (s, 19-CH₃), 2.13 (s, 21-OAc), 3.01 (t, J = 8, 17 α -H), 3.25 and 3.55 (ABq, J = 8.2, 18-CH₂), 4.24 (d, J = 6.5, 11 α -H), 4.53 (m, 3 β -H), 4.53 (s, 21-CH₂) ppm; m/e 330 (M-60).

The diacetate **8c** was prepared by dissolving **8b** in 5 weights each of pyridine and Ac₂O. Next day the product was ppt with ice-water and recrystallized from MeOH, m.p. 126-128°; λ_{max}^{KBT} 5.72 and 5.78 μ ; m/e 432.

 $3\alpha_2 21 - Dihydroxy - 11\beta_1 18 - oxido - 5\beta - pregnane 20 - one$ (8a). A soln of 203 mg of 8c in 2.6 ml MDC and 8.3 ml MeOH wastreated with a mixture of 1.2 ml cone HCl and 0.65 ml water. Afterstoring for 3 days at room temp. the mixture was worked up asfor preparation of 11a and the oily product chromatographed.Elution with petroleum ether-acetone 1:1 furnished 85 mg of 8a,mp. 145-151°, which after recrystallization from EtOAc-ether $melted at 148-152°: <math>\lambda_{max}^{EB} 2.94$ and 5.79 μ .

Acetylation of 8a with Ac₂O-pyridine afforded a compound identical with the starting 8c.

21 - Hydroxy - 11 β ,18 - oxido - 5 β - pregnane - 3,20 - dione acetate (5b). A soln of 100 mg of 8b in 4 ml acetone was treated at 0° with 0.5 ml of Jones reagent. After 5 min 1.5 ml MeOH was added, followed by 15 ml water. The soln was concentrated *in* vacuo at room temp. to a volume of 5 ml, and the crystalline dione 5b was collected and washed with water to afford 72 mg, m.p. 103-106°. Two recrystallizations from MDC-etherpetroleum ether raised the m.p. to 109-111°; $\lambda_{max}^{KBr} 5.70, 5.79$ and 5.88 μ ; δ 1.05 (s, 19-CH₃), 2.13 (s, 21-OAc), 3.01 (t, J = 8, 17 α -H), 3.30 and 3.60 (ABq, J = 8, 18-CH₂), 4.28 (d, J = 6.5, 11 α -H), 4.54 (s, 21-CH₂) ppm; *ml* e 328 (M-60). Acidic hydrolysis of 5b, as described for the preparation of 11a, afforded the 21-ol 5a. After chromatography (elution with 20% acetone in petroleum ether) the product was recrystallized from MDC-ether and melted at 153-155°; λ_{max}^{KBr} 2.88 and 5.85 μ ; m/e 346.

Acetylation of 5a with Ac₂O-pyridine as described gave 5b, identical with the sample described.

Hydrogenation of the ketoester **5b**. A suspension of 325 mg of **5b**, m.p. 100-104°, in 30 ml EtOH containing 0.3 ml AcOH, was hydrogenated for 96 h in a Parr apparatus at 20 psi at room temp. in the presence of 500 mg 5%Pd-C. The filtered soln was evaporated *in vacuo* to dryness, the residue taken up in MDC and washed with NaHCO₃aq. The organic soln was evaporated and the residue chromatographed: elution with 20% acetone in petroleum ether yielded at first unreacted **5b** (77 mg), then 3 β ,21 - dihydroxy - 11 β ,18 - oxido - 5 β - pregnane - 20 - one 21 - acetate **9** (146 mg), m.p. 146-148° (ether or MDC-petroleum ether); $\lambda_{\rm KBy}^{\rm KBy}$ 2.83, 5.75 and 5.82 μ ; δ 0.99 (s, 19-CH₃), 2.13 (s, 21-OAc), 3.21 and 3.50 (ABq, J = 8.2, 18-CH₂), 3.92 (m, 3 α -H), 4.00 (m, 17 α -H), 4.22 (d, J = 6.5, 11 α -H), 4.47 (s, 21-CH₂) ppm; *mle* 330 (M-60).

Further elution yielded 15 mg of the 3α -epimer 8b, m.p. 100-105° (ether).

Oxidation of 9 into 5b was carried out as described for the conversion of the 3α -isomer 8b into 5b: starting with 20 mg of 9, 12 mg of air-dried 5b was isolated, m.p. 104-107°, identical with authentic material.

21 - Hydroxy - 11 β ,18 - oxido - 5 α - pregnane - 20 - one acetate (13). A soln of 120 mg of 12 in 25 ml EtOAc was hydrogenated at atmospheric pressure for 2.5 h in the presence of 200 mg 5% Pd-C. The product was best purified by passage through a column of silica gel (elution with 5% acetone in petroleum ether) to furnish 87 mg of m.p. 132-135° (petroleum ether); $\lambda \frac{\text{Kib}}{\text{max}}$ 5.70 and 5.80 μ ; δ 0.87 (s, 19-CH₃), 2.16 (s, 21-OAc), 3.07 (t, J = 8.5, 17 α -H), 3.34 and 3.70 (ABq, J = 8.2, 18-CH₂), 4.41 (d, J = 6.5, 11 α -H), 4.63 (s, 21-CH₂) pm; m/e 314 (M-60).

Acknowledgement—The authors wish to thank Mrs. Sharon Linder for secretarial services, Mrs. Ronit Weininger for the NMR measurements and Mr. Shimon Hauptmann for recording the mass spectra.

REFERENCES

¹R. Neher, J. Endocr. 81, 25P (1975).

- ²S. Ulick, D. Marver, W. R. Adam and J. W. Funder, *Endo*crinology 104, 1352 (1979).
- ³J. W. Funder, J. Steroid Biochem. 11, 87 (1979).
- ⁴D. J. Morris, Endocrine Reviews 2, 234 (1981).
- ⁵J. Schmidlin, G. Anner, J. R. Billeter, K. Heusler, H. Ueberwasser, P. Wieland and A. Wettstein, *Helv. Chim. Acta* 40, 2291 (1957).
- ²²⁹¹ (1937). ⁶J. Schmidlin and A. Wettstein, *Ibid.* 43, 973 (1960).
- ⁷J. Schmidlin and A. Wettstein, *Ibid.* 43, 575 (1960).
- ⁸E. Kondo, T. Mitsugi and K. Tori, J. Am. Chem. Soc. 87, 4655
- (1965); U.S. pat. 3,780,026 (Dec. 18, 1973). ⁹L. J. Chinn, B. N. Desai and G. R. Lenz, J. Chem. Res. (S) 284
- (1978). ¹⁰O. Mancera, G. Rosenkranz and F. Sondheimer, J. Am. Chem.
- Soc. 77, 5669 (1955).
- ¹¹M. Harnik, Steroids 2, 485 (1963).
- ¹²Y. Lederman, R. Szpigielman, M. Bendcovsky, J. Herling and M. Harnik, Anal. Biochem. 51, 193 (1973).
- ¹³M. Harnik, Y. Lederman, R. Szpigielman and J. Herling, *Tetrahedron* 32, 1001 (1976).
- ¹⁴^aM. Harnik, Y. Aharonowitz and R. Lamed, *Tetrahedron* 38, 3173 (1982); ^brefs 13-18 contained therein.
- ¹⁵D. N. Kirk and B. W. Miller, J. Chem. Soc. Perkin I 2818 (1980).